World Organisation for Animal Health

Kyle Garver - - CANADA

WOAH Reference Laboratory Reports Activities 2024

This report has been submitted: 30 janvier 2025 22:49

LABORATORY INFORMATION

***Name of disease (or topic) for which you are a designated WOAH Reference Laboratory:**	Viral haemorrhagic septicaemia
***Address of laboratory:**	Pacific Biological Station, 3190 Hammond Bay Road, Nanaimo, British Columbia, Canada, V9T 6N7
***Tel:**	+1-250 756 73 40
***E-mail address:**	Kyle.Garver@dfo-mpo.gc.ca
Website:	https://profils-profiles.science.gc.ca/en/profile/kyle-garver
***Name (including Title) of Head of Laboratory (Responsible Official):**	Andrew Thomson (Regional Director of Science)
***Name (including Title and Position) of WOAH Reference Expert:**	Dr. Kyle Garver, Research Scientist
***Which of the following defines your laboratory? Check all that apply:**	Governmental

TOR1: DIAGNOSTIC METHODS

1. Did your laboratory perform diagnostic tests for the specified disease/topic for purposes such as disease diagnosis, screening of animals for export, surveillance, etc.? (Not for quality control, proficiency testing or staff training)

Yes

Diagnostic Test	Indicated in WOAH Manual (Yes/No)	Total number of test performed last year	
Indirect diagnostic tests		Nationally	Internationally
Direct diagnostic tests		Nationally	Internationally
RT-qPCR	Yes	415	

			8
RT-PCR	Yes	2	0
Virus Isolation	Yes	347	0

TOR2: REFERENCE MATERIAL

2. Did your laboratory produce or supply imported standard reference reagents officially recognised by WOAH?

No

3. Did your laboratory supply standard reference reagents (nonWOAH-approved) and/or other diagnostic reagents to WOAH Members?

Yes

Type of reagent available	Related diagnostic test	Produced/ provide	Amount supplied nationally (ml, mg)	Amount supplied internationally (ml, mg)	No. of recipient WOAH Member Countries	Country of recipients
Liquid Extraction controls- Artificial RNA transcript containing primer/probe binding sites spiked in EPC cell suspension	RT-qPCR (Garver et al 2011)	Produced	10 Aliquots (2.5mL)	0	1	CANADA,
Tissue Extraction - Naive kidney tissue spiked with artificial RNA transcript containing primer/probe binding sites	RT-qPCR (Garver et al 2011)	Produced	30 Aliquots (2250mg)	0	1	CANADA,
RT controls - Artificial RNA transcript	RT-qPCR (Garver et al 2011)	Produced	40 Aliquots (0.48mL)	0	1	CANADA,
qPCR controls – cDNA generated from Artificial RNA transcript	RT-qPCR (Garver et al 2011)	Produced	120 Aliquots (1.44mL)	0	1	CANADA,
Cell Lines	Cell culture	Provided	5 T75 flasks	0	1	CANADA,

4. Did your laboratory produce vaccines?

Not applicable

5. Did your laboratory supply vaccines to WOAH Members?

Not applicable

TOR3: NEW PROCEDURES

6. Did your laboratory develop new diagnostic methods for the designated pathogen or disease?

No

7. Did your laboratory validate diagnostic methods according to WOAH Standards for the designated pathogen or disease?

No

8. Did your laboratory develop new vaccines for the designated pathogen or disease?

No

9. Did your laboratory validate vaccines according to WOAH Standards for the designated pathogen or disease?

No

TOR4: DIAGNOSTIC TESTING FACILITIES

10. Did your laboratory carry out diagnostic testing for other WOAH Members?

No

11. Did your laboratory provide expert advice in technical consultancies on the request of an WOAH Member?

Yes

Name of the WOAH Member Country receiving a technical consultancy	Purpose	How the advice was provided
CANADA	advice concerning test capacity	remote
CANADA	Clinical disease symptoms	remote
KOREA (REP. OF)	Biosecurity requirements	remote
DENMARK	Diagnostic test recommendations	remote and in-person

TOR5: COLLABORATIVE SCIENTIFIC AND TECHNICAL STUDIES

12. Did your laboratory participate in international scientific studies in collaboration with WOAH Members other than the own?

Yes

Title of the study	Duration	Purpose of the study	Partners (Institutions)	WOAH Member Countries involved other than your country
Epidemiology of VHSV genotype IV	2022-2025	genetic diversity and distribution of VHSV genotype IV	Western Fisheries Research Center	UNITED STATES OF AMERICA
Test method review and comparison	2024-2025	Optimize diagnostic test methods	EURL for fish and crustacean diseases	DENMARK
Rapid sequencing method development	2024	Develop sequencing pipeline for rapid genotyping	Australian Centre for Disease Preparedness (ACDP) | CSIRO ACDP Fish Diseases Laboratory	AUSTRALIA

13. In exercising your activities, have you identified any regulatory research needs* relevant for WOAH?

No

TOR6: EPIZOOLOGICAL DATA

14. Did your Laboratory collect epidemiological data relevant to international disease control?

Yes

If the answer is yes, please provide details of the data collected:
Survey of wild and farmed fish populations for VHSV

15. Did your laboratory disseminate epidemiological data that had been processed and analysed?

Yes

If the answer is yes, please provide details of the data collected:
VHSV prevalence and genotype circulating withing wild AND cultured fish populations

16. What method of dissemination of information is most often used by your laboratory? (Indicate in the appropriate box the number by category and list the details in the box)

a) Articles published in peer-reviewed journals:

b) International conferences:

1
Australian Aquatic Animal Health Technical Forum and Skills Training Workshop - Invited speaker

c) National conferences:

d) Other (Provide website address or link to appropriate information):

TOR7: SCIENTIFIC AND TECHNICAL TRAINING

17. Did your laboratory provide scientific and technical training to laboratory personnel from other WOAH Members?

No

TOR8: QUALITY ASSURANCE

18. Does your laboratory have a Quality Management System?

Yes

Quality management system adopted	Certificate scan (PDF, JPG, PNG format)	
ISO/IEC 17025:2017	pdf	ASB_SOA_151008_FY23_v1_2023-07-31.pdf

19. Is your quality management system accredited?

Yes

Test for which your laboratory is accredited	Accreditation body
Reverse Transcription Quantitative PCR for Detection of Infectious Hematopoietic Necrosis Virus (IHNV)	Standards Council of Canada
Reverse Transcription Quantitative PCR for Detection of Viral Hemorrhagic Septicemia Virus (VHSV)	Standards Council of Canada
Reverse Transcription Quantitative PCR for Detection of Infectious Pancreatic Necrosis Virus (IPNV)	Standards Council of Canada
Isolation of Viral Agents (IPNV, IHNV, EHNV, SVCV, ISAV, SAV, and VHSV) from Finfish by cell culture	Standards Council of Canada
RT-qPCR Test method Protocol using TaqMan Universal PCR Master Mix for the detection of Infectious Salmon Anemia Virus	Standards Council of Canada
Histological Detection and Identification of Bivalve Mollusc Pathogens	Standards Council of Canada

20. Does your laboratory maintain a “biorisk management system” for the pathogen and the disease concerned?

Yes

Maintain laboratory compliance level 2 for in vitro facilities in accordance with the Canadian Biosafety Standard and the Containment Standards for Facilities Handling Aquatic Animal Pathogens

TOR9: SCIENTIFIC MEETINGS

21. Did your laboratory organise scientific meetings related to the pathogen in question on behalf of WOAH?

No

22. Did your laboratory participate in scientific meetings related to the pathogen in question on behalf of WOAH?

No

TOR10: NETWORK WITH WOAH REFERENCE LABORATORIES

23. Did your laboratory exchange information with other WOAH Reference Laboratories designated for the same pathogen or disease?

Yes

24. Do you network (collaborate or share information) with other WOAH Reference Laboratories designated for the same pathogen?

Yes

NETWORK/DISEASE	ROLE OF YOUR LABORATORY (PARTICIPANT, ORGANISER, ETC)	NO. PARTICIPANTS	PARTICIPATING WOAH REF. LABS

VHSV annual ref lab check-in	co-organizer	2	EURL and PBS-AAHL

25. Did you organise or participate in inter-laboratory proficiency tests with WOAH Reference Laboratories designated for the same pathogen during the past 2 years?

Yes

Purpose of the proficiency test:	Role of your Reference Laboratory (organiser/ participant)	No. participating Laboratories	Participating WOAH Ref. Labs/ organising WOAH Ref Lab
Interlaboratory proficiency test by EURL for Fish and Crustacean Diseases	participant	45	Participating WOAH reference laboratory for VHSV in Korea, Canada, and Denmark. Organized by reference laboratory in Denmark

26. Did your laboratory collaborate with other WOAH Reference Laboratories for the same disease on scientific research projects for the diagnosis or control of the pathogen of interest?

No

TOR11: OTHER INTERLABORATORY PROFICIENCY TESTING

27. Did your laboratory organise or participate in inter-laboratory proficiency tests with laboratories other than WOAH Reference Laboratories for the same pathogen during the past 2 years?

Yes

Purpose for inter-laboratory test comparisons1	Role of your reference laboratory (organizer/participant)	No. participating laboratories	Name of the test	WOAH Member Countries
Checking and certifying the performance of individual operators	organizer	3	RT-qPCR	CANADA,
Assess competency for diagnosis of fish diseases including IHN (Participate in the inter-laboratory PT from EU Reference Laboratory for fish and crustacean diseases)	participant	45	Virus isolation and RT-qPCR	AUSTRALIA, BOSNIA AND HERZEGOVINA, CANADA, CHILE, DENMARK, FAROE (ISLANDS), FINLAND, FRANCE, GERMANY, HUNGARY, ICELAND, ITALY, JAPAN, KOREA (REP. OF), LATVIA, NORWAY, SWEDEN, THE NETHERLANDS, UNITED STATES OF AMERICA,

TOR12: EXPERT CONSULTANTS

Kyle Garver - - CANADA

28. Did your laboratory place expert consultants at the disposal of WOAH?

Yes

Kind of consultancy	Location	Subject (facultative)
Review of Aquatic Animals Commission Report	remote	test recommendations

29. Additional comments regarding your report:

No