

WOAH Reference Laboratory Reports Activities 2024

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LABORATORY INFORMATION

*Name of disease (or topic) for which you are a designated WOAHO Reference Laboratory:	Infectious haematopoietic necrosis
*Address of laboratory:	Pacific Biological Station, 3190 Hammond Bay Road, Nanaimo, British Columbia, Canada, V9T 6N7
*Tel:	+1-250 756 73 40
*E-mail address:	Kyle.Garver@dfo-mpo.gc.ca
Website:	https://profils-profiles.science.gc.ca/en/profile/kyle-garver
*Name (including Title) of Head of Laboratory (Responsible Official):	Andrew Thomson (Regional Director of Science)
*Name (including Title and Position) of WOAHO Reference Expert:	Dr. Kyle Garver, Research Scientist
*Which of the following defines your laboratory? Check all that apply:	Governmental

TOR1: DIAGNOSTIC METHODS

1. Did your laboratory perform diagnostic tests for the specified disease/topic for purposes such as disease diagnosis, screening of animals for export, surveillance, etc.? (Not for quality control, proficiency testing or staff training)

Yes

Diagnostic Test	Indicated in WOAHO Manual (Yes/No)	Total number of test performed last year	
Indirect diagnostic tests		Nationally	Internationally
Direct diagnostic tests		Nationally	Internationally
RT-qPCR	Yes	183	

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			8
Virus Isolation	Yes	568	0

TOR2: REFERENCE MATERIAL

2. Did your laboratory produce or supply imported standard reference reagents officially recognised by WOA?

No

3. Did your laboratory supply standard reference reagents (nonWOAH-approved) and/or other diagnostic reagents to WOA Members?

Yes

Type of reagent available	Related diagnostic test	Produced/ provide	Amount supplied nationally (ml, mg)	Amount supplied internationally (ml, mg)	No. of recipient WOA Member Countries	Country of recipients
Tissue extraction control - Naive kidney tissue spiked with artificial RNA transcript containing primer/probe binding sites	RT-qPCR (Purcell et al. 2013)	Produced	15 aliquots (1125mg)	0	1	CANADA,
RT controls - Artificial RNA transcript	RT-qPCR (Purcell et al. 2013)	Produced	10 aliquots (0.12mL)	0	1	CANADA,
qPCR controls – cDNA generated from Artificial RNA transcript	RT-qPCR (Purcell et al. 2013)	Produced	40 aliquots (0.48mL)	0	1	CANADA,
Cell lines	Cell Culture	Provide	5 T75 flasks	0	1	CANADA,

4. Did your laboratory produce vaccines?

No

5. Did your laboratory supply vaccines to WOA Members?

No

TOR3: NEW PROCEDURES

6. Did your laboratory develop new diagnostic methods for the designated pathogen or disease?

No

7. Did your laboratory validate diagnostic methods according to WOA Standards for the designated pathogen or disease?

No

8. Did your laboratory develop new vaccines for the designated pathogen or disease?

No

9. Did your laboratory validate vaccines according to WOA Standards for the designated pathogen or disease?

No

TOR4: DIAGNOSTIC TESTING FACILITIES

10. Did your laboratory carry out diagnostic testing for other WOAHH Members?

No

11. Did your laboratory provide expert advice in technical consultancies on the request of an WOAHH Member?

Yes

Name of the WOAHH Member Country receiving a technical consultancy	Purpose	How the advice was provided
CANADA	Outbreak response simulation advice	remote
KOREA (REP. OF)	Test validation advice	remote
KOREA (REP. OF)	Biosecurity requirements	remote
DENMARK	Diagnostic methodology recommendations	remote and in-person
UNITED STATES OF AMERICA	Advice regarding detections and virulence potential	in-person

TOR5: COLLABORATIVE SCIENTIFIC AND TECHNICAL STUDIES

12. Did your laboratory participate in international scientific studies in collaboration with WOAHH Members other than the own?

Yes

Title of the study	Duration	Purpose of the study	Partners (Institutions)	WOAHH Member Countries involved other than your country
Epidemiology of IHN in the North Pacific	2018-2025	Genotype IHN	Western Fisheries Research Center	UNITED STATES OF AMERICA
Validation and comparison of test methods	2024-2026	Develop new and improved test methods	EURL for fish diseases and crustaceans	DENMARK
High throughput sequencing methodology development	2024	Optimize sequencing pipelines	Australian Centre for Disease Preparedness (ACDP) CSIRO ACDP Fish Diseases Laboratory	AUSTRALIA

13. In exercising your activities, have you identified any regulatory research needs* relevant for WOAHH?

Yes

Research need : 1

Please type the Research need: Validation of a modified diagnostic assay for IHN

Relevance for WOAHH Standard Setting,

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Relevance for the Code or Manual Manual,

Field Diagnostics,

Animal Category Aquatic,

Disease:

Infection with infectious haematopoietic necrosis virus

Kind of disease (Zoonosis, Transboundary diseases) Transboundary diseases,

If any, please specify relevance for Codes or Manual, chapter and title

(e.g. Terrestrial Manual Chapter 2.3.5 - Minimum requirements for aseptic production in vaccine manufacture)

Answer: infection with infectious haematopoietic necrosis virus

Notes:

Answer:

TOR6: EPIZOOLOGICAL DATA

14. Did your Laboratory collect epidemiological data relevant to international disease control?

Yes

If the answer is yes, please provide details of the data collected:

Surveillance of wild and farmed fish stocks for IHNV

15. Did your laboratory disseminate epidemiological data that had been processed and analysed?

Yes

If the answer is yes, please provide details of the data collected:

Nucleic acid and infectious virus detections in wild and cultured salmon stocks of Canada

16. What method of dissemination of information is most often used by your laboratory? (Indicate in the appropriate box the number by category and list the details in the box)

a) Articles published in peer-reviewed journals:

b) International conferences:

1

Australian Aquatic Animal Health Technical Forum and Skills Training Workshop

c) National conferences:

d) Other (Provide website address or link to appropriate information):

TOR7: SCIENTIFIC AND TECHNICAL TRAINING

17. Did your laboratory provide scientific and technical training to laboratory personnel from other WOA Members?

No

TOR8: QUALITY ASSURANCE

18. Does your laboratory have a Quality Management System?

Yes

Quality management system adopted	Certificate scan (PDF, JPG, PNG format)	
ISO/IEC 17025:2017	PDF	ASB_SOA_151008_FY23_v1_2023-07-31.pdf

19. Is your quality management system accredited?

Yes

Test for which your laboratory is accredited	Accreditation body
Reverse Transcription Quantitative PCR for Detection of Infectious Hematopoietic Necrosis Virus (IHNV)	Standards Council of Canada
Reverse Transcription Quantitative PCR for Detection of Viral Hemorrhagic Septicemia Virus (VHSV)	Standards Council of Canada
Reverse Transcription Quantitative PCR for Detection of Infectious Pancreatic Necrosis Virus (IPNV)	Standards Council of Canada
Isolation of Viral Agents (IPNV, IHNV, EHN, SVCV, ISAV, SAV, and VHSV) from Finfish by cell culture	Standards Council of Canada
RT-qPCR Test method Protocol using TaqMan Universal PCR Master Mix for the detection of Infectious Salmon Anemia Virus	Standards Council of Canada
Histological Detection and Identification of Bivalve Mollusc Pathogens	Standards Council of Canada

20. Does your laboratory maintain a "biorisk management system" for the pathogen and the disease concerned?

Yes

Maintain laboratory compliance level 2 for in vitro facilities in accordance with the Canadian Biosafety Standard and the Containment Standards for Facilities Handling Aquatic Animals Pathogens

TOR9: SCIENTIFIC MEETINGS

21. Did your laboratory organise scientific meetings related to the pathogen in question on behalf of WOAHP?

No

22. Did your laboratory participate in scientific meetings related to the pathogen in question on behalf of WOAHP?

No

TOR10: NETWORK WITH WOAHP REFERENCE LABORATORIES

23. Did your laboratory exchange information with other WOAHP Reference Laboratories designated for the same pathogen or disease?

No

24. Do you network (collaborate or share information) with other WOAHP Reference Laboratories designated for the same pathogen?

No

25. Did you organise or participate in inter-laboratory proficiency tests with WOAHP Reference Laboratories designated for the same pathogen during the past 2 years?

Yes

Purpose of the proficiency test:	Role of your Reference Laboratory (organiser/ participant)	No. participating Laboratories	Participating WOAHP Ref. Labs/ organising WOAHP Ref Lab
Interlaboratory proficiency test by the European Union Reference Laboratory for Fish and Crustacean Diseases	participant	45	China, Korea, and Canada/ EURL

26. Did your laboratory collaborate with other WOAHP Reference Laboratories for the same disease on scientific research projects for the diagnosis or control of the pathogen of interest?

No

TOR11: OTHER INTERLABORATORY PROFICIENCY TESTING

27. Did your laboratory organise or participate in inter-laboratory proficiency tests with laboratories other than WOAHP Reference Laboratories for the same pathogen during the past 2 years?

Yes

Purpose for inter-laboratory test comparisons ¹	Role of your reference laboratory (organizer/participant)	No. participating laboratories	Name of the test	WOAHP Member Countries
Checking and certifying the performance of individual operators	organizer	2	RNA finfish pathogen RT-qPCR testing	CANADA,
Assess competency for diagnosis of fish diseases			Inter-laboratory Proficiency Test 2024 for identification	

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including IHN (Participate
in the inter-laboratory PT
from EU Reference
Laboratory for fish and
crustacean diseases)

participant

45

and titration of
VHSV,IHNV,EHNV (fish DENMARK,
ranaviruses), SVCV and IPNV
(PT1) and identification of
CyHV-3 (KHV), SAV and ISAV
(PT-2)

TOR12: EXPERT CONSULTANTS

28. Did your laboratory place expert consultants at the disposal of WOA?

Yes

Kind of consultancy	Location	Subject (facultative)
Review of WOA Aquatic Manual	remote	Validation
Responding to technical and training queries	remote	Advice and training opportunities
Review of WOA Standards	remote	Aquatic animal commission report
Participation in WOA surveys	remote	fish health research priorities

29. Additional comments regarding your report:

No