

# WOAH Reference Laboratory Reports Activities 2024

This report has been submitted: 3 mars 2025 00:59

# LABORATORY INFORMATION

*Name of disease (or topic) for which you are a designated WOAH Reference Laboratory:	Antimicrobial resistance	
*Address of laboratory:	Animal and Plant Health Agency, New Haw, Addlestone Surrey KT15 3NB Weybridge UNITED KINGDOM	
*Tel:	+44-20 8026 2578	
*E-mail address:	Christopher.Teale@apha.gov.uk	
Website:	http://apha.defra.gov.uk/apha-scientific/index.htm	
*Name (including Title) of Head of Laboratory (Responsible Official):	Dr J Stewart, Chief Executive, Animal and Plant Health Agency	
*Name (including Title and Position) of WOAH Reference Expert:	Dr Christopher Teale MRCVS, Head of Antimicrobial Resistance	
*Which of the following defines your laboratory? Check all that apply:	Veterinary Surveillance Governmental Research agency	

# **TOR1: DIAGNOSTIC METHODS**

1. Did your laboratory perform diagnostic tests for the specified disease/topic for purposes such as disease diagnosis, screening of animals for export, surveillance, etc.? (Not for quality control, proficiency testing or staff training)

Diagnostic Test	Indicated in WOAH Manual (Yes/No)	Total number of test performed last year	
Indirect diagnostic tests		Nationally	Internationally
No indirect tests are routinely performed.	No	0	0
Direct diagnostic tests		Nationally	Internationally
Disc diffusion susceptibility tests	Yes	7793	0

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Determination of minimum inhibitory concentration	Yes	13850	0
Whole genome sequencing	Yes	104	0

## **TOR2: REFERENCE MATERIAL**

2. Did your laboratory produce or supply imported standard reference reagents officially recognised by WOAH?

No

3. Did your laboratory supply standard reference reagents (nonWOAH-approved) and/or other diagnostic reagents to WOAH Members? No

4. Did your laboratory produce vaccines?

Not applicable

5. Did your laboratory supply vaccines to WOAH Members?

Not applicable

# **TOR3: NEW PROCEDURES**

6. Did your laboratory develop new diagnostic methods for the designated pathogen or disease?

No

7. Did your laboratory validate diagnostic methods according to WOAH Standards for the designated pathogen or disease? No

8. Did your laboratory develop new vaccines for the designated pathogen or disease?

No

9. Did your laboratory validate vaccines according to WOAH Standards for the designated pathogen or disease?

No

# **TOR4: DIAGNOSTIC TESTING FACILITIES**

10. Did your laboratory carry out diagnostic testing for other WOAH Members?

No

11. Did your laboratory provide expert advice in technical consultancies on the request of an WOAH Member?

No

# **TOR5: COLLABORATIVE SCIENTIFIC AND TECHNICAL STUDIES**

12. Did your laboratory participate in international scientific studies in collaboration with WOAH Members other than the own? Yes

Title of the study	Duration	Purpose of the study	Partners (Institutions)	WOAH Member Countries involved other than your country
AMR surveillance at live bird markets in Bangladesh	2 years	Determine prevalence of AMR	Central Disease Investigation Laboratory, Dhaka., Bangladesh	BANGLADESH

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MDR non-typhoidal Salmonella in migratory birds	4 years	Determine prevalence and AMR of non-typhoidal Salmonella in wild birds	Bangladesh Livestock Research Institute	BANGLADESH
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13. In exercising your activities, have you identified any regulatory research needs\* relevant for WOAH?

No

# **TOR6: EPIZOOLOGICAL DATA**

14. Did your Laboratory collect epidemiological data relevant to international disease control?

Yes

If the answer is yes, please provide details of the data collected:

Collated AMR data was published at https://www.gov.uk/government/publications/veterinary-antimicrobial-resistance-and-sales-surveillance-2023

15. Did your laboratory disseminate epidemiological data that had been processed and analysed?

Yes

If the answer is yes, please provide details of the data collected:

Prevalence of AMR in zoonotic and commensal bacteria in healthy animals; prevalence of AMR in bacterial veterinary pathogens.

16. What method of dissemination of information is most often used by your laboratory? (Indicate in the appropriate box the number by category and list the details in the box)

a) Articles published in peer-reviewed journals:

#### 16

Kirchner M; Nunez-Garcia J; Duggett N; Gosling RJ; Anjum MF (2024) Use of transcriptomics and genomics to assess the effect of disinfectant exposure on the survival and resistance of Escherichia coli O157:H7, a human pathogen. Frontiers in Microbiology 15 https://doi.org/10.3389/fmicb.2024.1477683

Eric J Threlfall, Roberto La Ragione, Rohini Manuel, Chris Teale, Paul Cook, Bobby Kainth, Henry Nicholls, Kathryn Callaghan. Advisory Committee on the Microbiological Safety of Food recommendations for standardisation of antimicrobial resistance terminology research comment published in Veterinary Record November 2024, 415-417 Advisory Committee on the Microbiological Safety of Food recommendations for standardisation of antimicrobial resistance terminology

Sati NM; Card RM; Barco L; Muhammad M; Luka PD; Chisnall T; Fagbamila IO; Cento G; Nnadi NE; Kankya C; Rwego IB; Ikwap K; Mugisha L; Erume J; Mwiine FN (2024) Antimicrobial Resistance and Phylogenetic Relatedness of Salmonella Serovars in Indigenous Poultry and Their Drinking Water Sources in North Central Nigeria. Microorganisms 12 (8) https://doi.org/10.3390/microorganisms 12081529



Read DS; Gweon HS; Bowes MJ; Anjum MF; Crook DW; Chau KK; Shaw LP; Hubbard A; Abuoun M;

Tipper HJ; Hoosdally SJ; Bailey MJ; Walker AS; Stoesser N (2024) Dissemination and persistence of antimicrobial resistance (AMR) along the wastewater-river continuum. Water Research 264 https://doi.org/10.1016/j.watres.2024.122204

Gütgemann F, Heuvelink A, Müller A, Churin Y, Buter R, Jung A, Feberwee A, Wiegel J, Kumm F, Braun AS, Yue M, Soriano-Vargas E, Swanepoel S, Botteldoorn N, Kirchner M, Kehrenberg C. Recommendation of a standardized broth microdilution method for antimicrobial susceptibility testing of Avibacterium paragallinarum and resistance monitoring. J Clin Microbiol. 2024 Feb 16:e0101123. https://doi.org/10.1128/jcm.01011-23

Muna F Anjum, Nicholas Duggett, Ewart Sheldon, Meenaxi Sharma, Richard P Smith, Chris J Teale. Genomics to detect transmission of livestock-associated methicillin-resistant Staphylococcus aureus from UK pigs in abattoirs during slaughter Journal of Antimicrobial Chemotherapy, dkae052, https://doi.org/10.1093/jac/dkae052

Shafiq Rheman, Sabrina Hossain, Md Samun Sarker, Farhana Akter, Laura Khor, Han Ming Gan, Andy Powell, Roderick M. Card, Yaovi Mahuton Gildas Hounmanou, Anders Dalsgaard, Chadag Vishnumurthy Mohan, Zamila Bueaza Bupasha, Mohammed A. Samad, David W. Verner-Jeffreys, Jérôme Delamare-Deboutteville Nanopore sequencing for identification and characterization of Antimicrobial-Resistant Escherichia coli and Salmonella spp. from tilapia and shrimp sold at wet markets in Dhaka, Bangladesh Frontiers of Microbiology, https://doi:10.3389/fmicb.2024.1329620

Jennifer Mattock, Marie Anne Chattaway, Hassan Hartman, Timothy J. Dallman, Anthony M. Smith, Karen Keddy, Liljana Petrovska, Emma J. Manners, Sanelisiwe T. Duze, Shannon Smouse, Nomsa Tau, Ruth Timme, Dave J. Baker, Alison E. Mather, John Wain, and Gemma C. Langridge A One Health Perspective on Salmonella enterica Serovar Infantis, an Emerging Human Multidrug-Resistant Pathogen Emerging Infectious Diseases Journal, Volume 30, Number 4—April 2024 https://wwwnc.cdc.gov/eid/article/30/4/23-1031\_article

Duggett N; Abuoun M; Stubberfield E; Turner O; Randall L; Horton R; Nunez-Garcia J; Gates D; Chanter J; Teale C; Anjum MF (2023) Genomic surveillance of extended-spectrum cephalosporin-resistant Escherichia coli isolated from poultry in the UK from 2016 to 2020. Frontiers in Microbiology 14 https://doi.org/10.3389/fmicb.2023.1335173

Huong LQ; Chisnall T; Rodgers JD; Cawthraw SA; Card RM (2024) Prevalence, antibiotic resistance, and genomic characterisation of Campylobacter spp. in retail chicken in Hanoi, Vietnam. Microbial Genomics 10 (1) https://doi.org/10.1099/mgen.0.001190

Stubberfield E; Abuoun M; Card RM; Welchman D; Anjum MF (2024) Molecular characterization of antimicrobial resistance in Brachyspira species isolated from UK chickens: Identification of novel variants of pleuromutilin and beta-lactam resistance genes. Veterinary Microbiology 290

https://doi.org/10.1016/j.vetmic.2024.109992

Olorunleke SO; Kirchner M; Duggett N; Stevens K; Chah KF; Nwanta JA; Brunton LA; Anjum MF (2024) Rapid detection and molecular epidemiology of β-lactamase producing Enterobacteriaceae isolated from food animals and in-contact humans in Nigeria. PLoS ONE 19 (4 April)

#### https://doi.org/10.1371/journal.pone.0289190

Gand M; Navickaite I; Bartsch LJ; Grützke J; Overballe-Petersen S; Rasmussen A; Otani S; Michelacci V; Matamoros BR; González-Zorn B; Brouwer MSM; Di Marcantonio L; Bloemen B; Vanneste K; Roosens NHCJ; Abuoun M; De Keersmaecker SCJ (2024) Towards facilitated interpretation of shotgun metagenomics long-read sequencing data analyzed with KMA for the detection of bacterial pathogens and their antimicrobial resistance genes. Frontiers in Microbiology 15 https://doi.org/10.3389/fmich\_2024\_1336532

https://doi.org/10.3389/fmicb.2024.1336532

Davies TJ; Swann J; Sheppard AE; Pickford H; Lipworth S; Abuoun M; Ellington MJ; Fowler PW; Hopkins S; Hopkins KL; Crook DW; Peto TEA; Anjum MF; Walker AS; Stoesser N (2023) Discordance between different bioinformatic methods for identifying resistance genes from short read genomic data, with a focus on Escherichia coli Microbial Genomics 9 (12) https://doi.org/10.1099/mgen.0.001151



Deza-Cruz I; Vilar MJ; Velasova M; Abuoun M; Anjum MF; Smith RP (2023) Antimicrobial resistance of Escherichia coli in the UK: comparison of single vs. pooled samples from healthy pigs. Letters in Applied Microbiology 76 (11) https://doi.org/10.1093/lambio/ovad123

b) International conferences:

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Attended the second annual congress of FAO designated Reference Centres for AMR in Rome (19-20 September 2024). Attended the 8th World One Health Congress (WOHC) in Cape Town, South Africa (20 to 23 September 2024).

c) National conferences:

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Attended the launch meeting for the AMR in Agrifood Systems Transdisciplinary (AMAST) Network. Wellcome Sanger conference 'Antimicrobial Resistance – Genomes, Big Data and Emerging Technologies', Cambridge, UK.

d) Other (Provide website address or link to appropriate information):

#### 3

APHA Science Blogs for World AMR Awareness Week, themed on "Combatting antimicrobial resistance through global training", "Filling gaps in our understanding of antimicrobial resistance to safeguard animal and human health" and "A visit of a lifetime: Our experience with APHA's UK FAO Reference Centre for AMR"

https://aphascience.blog.gov.uk/category/amr/

## **TOR7: SCIENTIFIC AND TECHNICAL TRAINING**

17. Did your laboratory provide scientific and technical training to laboratory personnel from other WOAH Members?

No

# **TOR8: QUALITY ASSURANCE**

18. Does your laboratory have a Quality Management System?

Quality management system adopted	Certificate scan (PDF, JPG, PNG format)	
ISO17025	PDF	17025 certificate.pdf

19. Is your quality management system accredited?

Yes

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Test for which your laboratory is accredited	Accreditation body
Disc diffusion test	UKAS

20. Does your laboratory maintain a "biorisk management system" for the pathogen and the disease concerned?

Yes

Advisory Committee on Dangerous Pathogens

### **TOR9: SCIENTIFIC MEETINGS**

21. Did your laboratory organise scientific meetings related to the pathogen in question on behalf of WOAH?

No

22. Did your laboratory participate in scientific meetings related to the pathogen in question on behalf of WOAH?

# **TOR10: NETWORK WITH WOAH REFERENCE LABORATORIES**

23. Did your laboratory exchange information with other WOAH Reference Laboratories designated for the same pathogen or disease?

Not applicable (only WOAH Reference Laboratory designated for the disease

24. Do you network (collaborate or share information) with other WOAH Reference Laboratories designated for the same pathogen?

Not applicable (only WOAH Reference Laboratory designated for the disease

25. Did you organise or participate in inter-laboratory proficiency tests with WOAH Reference Laboratories designated for the same pathogen during the past 2 years?

Not applicable (Only WOAH Reference Laboratory designated for the disease) *Not applicable.* 

26. Did your laboratory collaborate with other WOAH Reference Laboratories for the same disease on scientific research projects for the diagnosis or control of the pathogen of interest?

Not applicable (only WOAH Reference Laboratory designated for the disease

# TOR11: OTHER INTERLABORATORY PROFICIENCY TESTING

27. Did your laboratory organise or participate in inter-laboratory proficiency tests with laboratories other than WOAH Reference Laboratories for the same pathogen during the past 2 years?

Yes

Purpose for inter-laboratory test comparisons1	Role of your reference laboratory (organizer/participant)	No. participating laboratories	Name of the test	WOAH Member Countries
External Quality Assurance of AMR testing in Salmonella, Campylobacter, Enterococci and E. coli	Organiser	14	Broth Dilution MIC	



# **TOR12: EXPERT CONSULTANTS**

28. Did your laboratory place expert consultants at the disposal of WOAH?

Yes

Kind of consultancy	Location	Subject (facultative)
Electronic Expert Group on AMR	Virtual Meetings	AMR.

29. Additional comments regarding your report:

No